

## Role of oxidative stress in the pathogenesis of hypertension

Yosit Ponce Gutiérrez<sup>a</sup>✉, MD; Arik Ponce Gutiérrez<sup>a</sup>, MD; Arnaldo Rodríguez León<sup>b</sup>, MD, MSc; and Katherin Cabrera García<sup>a</sup>, MD

<sup>a</sup> Juan B. Contreras Fowler Polyclinic. Ranchuelo, Villa Clara, Cuba.

<sup>b</sup> Dr. Celestino Hernández Robau University Hospital. Santa Clara, Villa Clara, Cuba.

*Este artículo también está disponible en español*

### ARTICLE INFORMATION

Received: October 14, 2013  
Accepted: November 5, 2013

### Competing interests

The authors declare no competing interests

### Acronyms

**ACE:** angiotensin-converting enzyme  
**DNA:** deoxyribonucleic acid  
**eNOS:** endothelial NOS  
**HT:** hypertension  
**NADPH:** nicotinamide adenine dinucleotide phosphate  
**NO:** nitric oxide  
**NOS:** nitric oxide synthase  
**OS:** oxidative stress  
**ROS:** reactive oxygen species  
**SOD:** superoxide dismutase  
**VSMC:** vascular smooth muscle cells

### On-Line Versions:

[Spanish](#) - [English](#)

✉ Y Ponce Gutiérrez  
Camilo Cienfuegos N° 63,  
e/ Carmen Rivero y Federico Escobar.  
Ranchuelo, Villa Clara, Cuba.  
E-mail address:  
[froilanponce@capiro.vcl.sld.cu](mailto:froilanponce@capiro.vcl.sld.cu)

### ABSTRACT

The increased production of reactive oxygen species has been involved in several chronic diseases, including hypertension. Oxidative stress is, in turn, cause and consequence of this hypertension. The enzyme NADPH oxidase is the major source of reactive species of cardiovascular, renal and neural oxygen. Oxidative stress is associated with endothelial dysfunction, inflammation, hypertrophy, apoptosis, cell migration, fibrosis and angiogenesis; important processes involved in vascular remodeling of hypertension. Despite the large amount of data that involve oxidative stress as a causative factor of experimental hypertension, results in humans are less conclusive. The aim of this review is to describe the role of oxidative stress in the pathophysiology of hypertension. A better understanding of these mechanisms will allow a more comprehensive behavior to this common disease.

**Key words:** Oxidative stress, Reactive oxygen species, Hypertension

### *Papel del estrés oxidativo en la patogénesis de la hipertensión arterial*

### RESUMEN

La producción aumentada de las especies reactivas de oxígeno ha sido implicada con varias enfermedades crónicas, incluida la hipertensión arterial. El estrés oxidativo es, a su vez, causa y consecuencia de esta hipertensión. La mayor fuente de especies reactivas de oxígeno cardiovascular, renal y neural es la enzima NADPH oxidasa. El estrés oxidativo se relaciona con disfunción endotelial, inflamación, hipertrofia, apoptosis, migración celular, fibrosis y angiogénesis; procesos importantes involucrados en la remodelación vascular de la hipertensión arterial. A pesar de la gran cantidad de datos que implican al estrés oxidativo como un factor causante de la hipertensión experimental, los resultados en humanos son menos conclusivos. El objetivo de esta revisión bibliográfica es describir el papel del estrés oxidativo en la fisiopatología de la hipertensión arterial. La mejor comprensión de estos mecanismos permitirá establecer una conducta más integral ante esta frecuente enfermedad.

**Palabras clave:** Estrés oxidativo, Especies reactivas del oxígeno, Hipertensión arterial

### INTRODUCTION

Hypertension (HT) is a chronic disease in which blood pressure (BP) shows le-

vels of 140/90 mmHg or higher. Worldwide, nearly one billion people suffer from HT, and it is estimated that this figure could increase to 1.5 billion by 2025. The exact genesis is unknown, since only about 5 % of hypertensive patients show a precise cause of this condition<sup>1</sup>.

At molecular level, numerous factors have been involved in the pathophysiology of HT, for example, the activation of the renin-angiotensin-aldosterone system, inflammation, aberrant G protein-coupled receptor signaling and endothelial dysfunction. Common to these factors is long-term oxidative stress, which causes overproduction of vascular reactive oxygen species (ROS), decrease in nitric oxide bioavailability and a reduction of antioxidant capacity<sup>2</sup>. In the vascular system, the ROS have a physiological role in the control of endothelial function and vascular tone, and an important pathophysiological role in inflammation, hypertrophy, proliferation, apoptosis, migration, fibrosis and angiogenesis. These are important factors in vascular remodeling and endothelial dysfunction associated with HT<sup>3</sup>.

Almost all experimental models of HT show some forms of oxidative excess. As the inhibition of ROS-generating enzymes, the antioxidants and ROS scavengers reduce blood pressure, whereas pro-oxidants increase blood pressure, it has been suggested that ROS are causally associated with hypertension, at least in animal models<sup>4</sup>. Although a large amount of data supports the role of oxidative stress in experimental HT, evidence in human HT is weak. A better understanding of these mechanisms will provide a deeper knowledge to ensure better prevention, control and the emergence of new therapeutic targets and novel treatments for this disease.

**Oxidative stress (OS)** is a state of the cell in which redox homeostasis is altered, that is, there is an imbalance between pro-oxidant and antioxidants<sup>5</sup>.

**Free radicals** are molecules whose atomic structure has an unpaired electron in the outer orbital, which gives them a special con-

figuration of great instability. This makes it very unstable, extremely reactive and short-lived, with a huge capacity for combining nonspecifically in most cases, as well as with the diversity of molecules of the cell structure: carbohydrates, lipids, proteins and nucleic acids<sup>5</sup>.

**Reactive oxygen species** are a variety of free radicals formed from O<sub>2</sub> (**Table**). ROS are inactivated by enzymatic or entrapment mechanisms<sup>5</sup>.

### HARMFUL EFFECT OF ROS ON BIOMOLECULES

Cellular damage caused by ROS occurs on different biomolecules.

#### Lipids

This is where the greatest damage occurs, in a process called lipid peroxidation, which affects structures that are rich in polyunsaturated fatty acids, since the permeability of the cell membrane is disrupted, so that edema and cell death follows. Lipid peroxidation or oxidative rancidity is a form of tissue damage which can be triggered by oxygen, singlet oxygen, hydrogen peroxide and hydroxyl radical. Unsaturated fatty acids are essential components of cell membranes, so they are believed to be important for its normal functioning; however, they are vulnerable to the oxidative attack initiated by ROS<sup>6</sup>.

The factors that influence the degree of the lipid peroxidation are:

- The qualitative and quantitative nature of the trigger agent.
- The polyunsaturated fatty acids of the membrane and their accessibility.
- Oxygen tension.

**Table.** Examples of ROS.

O <sub>2</sub> <sup>-</sup> : Superoxide	<sup>1</sup> O <sub>2</sub> : Singlet oxygen	ROO <sup>-</sup> : Peroxyl
OH: Hydroxyl radical	RO <sup>-</sup> : Alkoxy	ROOH: Organic hydroperoxide
H <sub>2</sub> O <sub>2</sub> : Hydrogen peroxide <sup>Ω</sup>	HOCl: Hypochlorous acid	ONOO <sup>-</sup> : Peroxynitrite

<sup>Ω</sup> It is not strictly a free radical, but is included as such for its ability to generate OH in the presence of metals (iron, copper). This process is called Fenton reaction.

- The presence of iron.
- The antioxidants in the cell (beta-carotene, alpha-tocopherol, glutathione).
- The activation of enzymes that may terminate the chain reaction, such as glutathione peroxidase.

Once started, the process takes the form of a “cascade” with ROS production leading to the formation of organic peroxides and other products from unsaturated fatty acids. Once formed, these ROS are responsible for the cytotoxic effects<sup>6</sup>.

### Proteins

There is the oxidation of a group of amino acids such as phenylalanine, tyrosine, histidine, and methionine; there are also peptide chains crosslinks, and finally, there is the formation of carbonyl groups<sup>5</sup>.

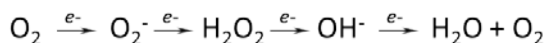
### Deoxyribonucleic acid (DNA)

There are mutations and carcinogenesis, loss of expression or synthesis of a protein due to damage to a specific gene, oxidative modifications of bases, deletions, fragmentation, stable DNA-protein interactions, chromosomal rearrangements and DNA cytosine demethylation that activates genes. The damage may occur due to alterations (inactivation/loss of some tumor suppressor genes that may lead to the initiation and progression of carcinogenesis). The tumor suppressor genes can be modified by a simple change in a critical base of the DNA sequence<sup>5</sup>.

### VASCULAR GENERATION OF ROS

ROS are produced as intermediates in redox reactions forming H<sub>2</sub>O and O<sub>2</sub>.

The sequential univalent reduction of O<sub>2</sub> is<sup>7,8</sup>:



In the ROS generated in vascular cells, O<sub>2</sub><sup>-</sup> (superoxide anion), and H<sub>2</sub>O<sub>2</sub> (hydrogen peroxide) seem to be particularly important. In biological systems, the O<sub>2</sub><sup>-</sup> is short-lived due to its rapid reduction to H<sub>2</sub>O<sub>2</sub> by the action of superoxide dismutase (SOD), of which three isoforms have been characterized in mammals:

copper/zinc SOD (SOD1), mitochondrial SOD (SOD2), and extracellular SOD (SOD3)<sup>9</sup>. The charge on the superoxide anion makes it difficult to cross the cellular membranes, except possibly through ion channels. H<sub>2</sub>O<sub>2</sub> has a longer lifespan than O<sub>2</sub><sup>-</sup>, is relatively stable and is easily diffusible within and between cells. The distinct chemical properties between O<sub>2</sub><sup>-</sup> and H<sub>2</sub>O<sub>2</sub> and their different sites of distribution suggest that different species of ROS activate diverse signaling pathways, which lead to divergent, and potentially opposing, biological responses<sup>10</sup>.

All vascular cell types produce ROS, including endothelial, smooth muscle, adventitial fibroblasts and perivascular adipocytes, and phagocyte cells (neutrophils, eosinophils, monocytes and macrophages). These ROS may be formed by many enzymes, including xanthine oxidoreductase, uncoupled nitric oxide synthase, mitochondrial respiratory enzymes and nicotinamide adenine dinucleotide phosphate (NADPH) oxidase. Of these, mitochondrial enzymes and NADPH oxidase seem to be particularly important in hypertension<sup>11-14</sup>.

### PATHOPHYSIOLOGY OF HT ASSOCIATED WITH OXIDATIVE STRESS

#### Endothelial dysfunction

Endothelial dysfunction has been involved in the pathophysiology of different forms of cardiovascular disease, including HT. This dysfunction can be defined as a deterioration of the actions of the endothelium in vasodilation, increased proinflammatory states and increased prothrombotic activity. These phenomena lead to a state of vascular inflammation that may be partially mediated by the ROS formed by activated mononuclear cells<sup>15</sup>.

#### Vascular OS and HT

The OS is a common mechanism of injury in many types of hypertensive disease processes, and occurs when there is an imbalance between ROS generation and antioxidant defense systems of the body. The ROS family comprises many molecules that have divergent effects on cell function. Indeed, many of these actions are associated with pathological changes observed in cardiovascular disease. ROS effects are mediated by

the redox-sensitive regulation of multiple signaling molecules and second messengers<sup>16-18</sup>. Several studies have shown excessive quantities of ROS in patients with essential HT and in several hypertensive animal models<sup>19-21</sup>; these patients and experimental animals have an inadequate antioxidant state<sup>22</sup>, which adds more evidence to the idea that oxidative stress may be involved in essential HT<sup>23</sup>. A strong relationship between BP and some parameters related to OS was recently demonstrated<sup>24</sup>. Other experimental studies showed that mice with genetic defects in ROS generating enzymes show lower values of BP compared to wild mice<sup>25,26</sup>. Furthermore, in cultured vascular smooth muscle cells of arteries from hypertensive rats and humans, ROS production is increased, the redox-dependent signaling is amplified and antioxidant activity is reduced<sup>27</sup>. The beneficial effects of classical antihypertensive agents such as beta-blockers, angiotensin-converting enzyme (ACE) inhibitors, angiotensin II receptor blockers, and calcium-channel blockers are mediated in part by a decrease in vascular OS<sup>28,29</sup>.

### The sources of ROS in the vascular wall

There is a variety of enzymatic and non-enzymatic sources of ROS in blood vessels. NADPH oxidase is the best characterized. Furthermore, various other enzymes may contribute to the generation of ROS, such as xanthine oxidase, nitric oxide synthase (NOS) and the mitochondrial enzymes.

### NADPH Oxidase

It is the primary biochemical source of ROS in the vasculature, particularly the  $O_2^-$ . It is also important in the kidney; therefore, it plays an important role in renal dysfunction and vascular damage in pathologic conditions<sup>30</sup>. This system catalyzes the reduction of molecular  $O_2$  by NADPH, which serves as an electron donor in the generation of  $O_2^-$ . NADPH oxidase is a positive regulator in HT by humoral and signaling mechanisms. Angiotensin II is the stimulus more commonly studied in the upregulation of NADPH oxidase, but endothelin-1 (ET-1) and urotensin II may also participate in the activation of NADPH oxidase, which leads to an increase in ROS. The best known effect of the  $O_2^-$  generated by NADPH oxidase is the inactivation of nitric oxide (NO) in a reaction that forms peroxynitrite, which damages the endothelial

vasodilation and uncouples endothelial NOS (eNOS), which leads to additional production of  $O_2^-$ <sup>25,31</sup>. In the vasculature, the activation of NADPH oxidase has been strongly associated with HT<sup>32</sup>.

### Uncoupling of eNOS

The primary function of eNOS is the production of the NO that regulates vasodilatation. However, the deficiency or oxidation of L-arginine and tetrahydrobiopterin ( $BH_4$ ), which are two essential cofactors in the functioning of eNOS, are associated with the uncoupling of the L-arginine-NO pathway, which results in a decrease in NO formation and an increase in the generation of  $O_2^-$ , mediated by the uncoupled eNOS.

The NADPH oxidase is the initial source of ROS. The  $O_2^-$  is combined with the NO, formed by eNOS, to form peroxynitrite<sup>33</sup>, which in turn oxidizes and destabilizes eNOS to produce additional  $O_2^-$ <sup>34</sup>. The  $O_2^-$  also leads to the oxidation of  $BH_4$ , which promotes eNOS uncoupling and a larger production of ROS.

### Xanthine oxidase

Xanthine oxidase is also a major source of ROS in the vascular endothelium<sup>35</sup>. It catalyzes the last two steps of purine metabolism. During this process, the  $O_2$  is reduced to  $O_2^-$ . There is evidence that suggests the involvement of this enzyme in HT. Hypertensive rats have shown high levels of endothelial xanthine oxidase and increased production of ROS, which are associated with increased arteriolar tone<sup>30</sup>. In addition to the effects on vasculature, this xanthine may have a role in target organ damage in hypertensive patients<sup>36</sup>.

### Mitochondria

Mitochondria are the biggest source and target of ROS. Some of the  $O_2^-$  that is produced in the intermembrane space may move to the cytoplasm<sup>37</sup>. Ubiquinone or coenzyme Q produces  $O_2^-$  when partially reduced (in form of semiquinone), and an antioxidant when it is fully reduced<sup>38</sup>. Complex I produces most of the  $O_2^-$  that is generated in mammalian's mitochondria. Complex II and IV are not normally significant sites of ROS production. A moderate uncoupling of complex I is very effective in reducing the production of  $O_2^-$  and it has been reported that there is a de-

crease in antioxidant enzyme activity in hypertensive patients<sup>39</sup>.

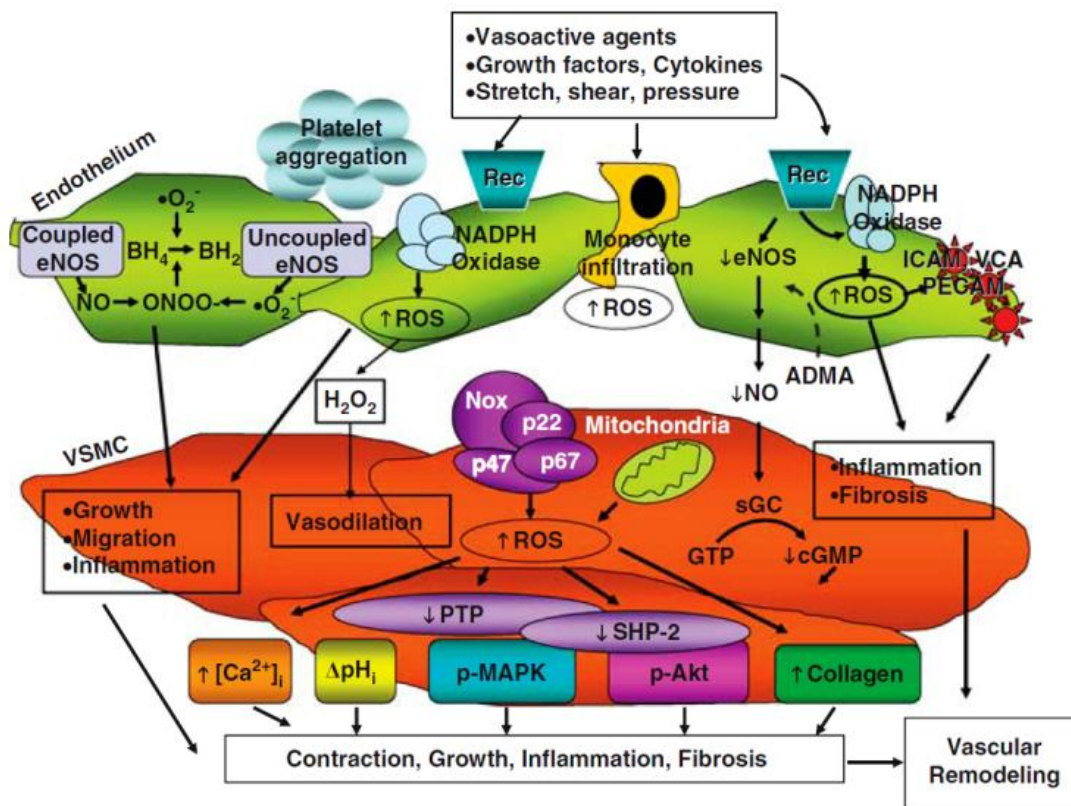
**Role of the components of the vascular wall**

The endothelium is sensitive to mechanical and hormonal changes; in response, it releases agents that regulate motor function. There is no doubt that the endothelium has a protective and regulatory role through the generation of vasorelaxant substances. Under pathological circumstances, the endothelium produces vasoconstrictors such as ET-1, angiotensin II, urotensin II, superoxide anions, vasoconstrictor pros-

taglandins and thromboxane A<sub>2</sub>, which may be released and contribute to the paradoxical vasoconstrictor effects.

The vascular smooth muscle cells (VSMC) are not only involved in the short-term regulation of blood vessel diameter, and therefore in the regulation of BP, but are also involved in long-term adaptation through structural remodeling. The ROS mediate many of these pathophysiological processes (**Figure**).

The adventitia may contribute to HT either by reducing the bioavailability of NO or by participating in vascular remodeling mediated by ROS.



**Figure.** Vascular remodeling induced by oxidative stress in hypertension. Activation of ROS-generating enzymes, such as NADPH oxidase, simultaneous uncoupling of NOS and mitochondrial enzymes in endothelial and vascular smooth muscle cells results in decreased NO production and increased generation of O<sub>2</sub><sup>-</sup> and H<sub>2</sub>O<sub>2</sub>, which in turn influence redox-sensitive signaling molecules including MAPKs, PTPs, ion channels, transcription factors that induce the expression of pro-inflammatory adhesion molecules such as intercellular adhesion molecules (ICAM), vascular cell adhesion molecules (VCAM), platelet endothelial cell adhesion molecule (PECAM). These processes lead to vascular growth, fibrosis, contraction/dilation, inflammation and platelet aggregation, which underlie vascular damage and structural remodeling in HT and other cardiovascular diseases.

**Legend:** ADMA (asymmetric dimethylarginine); BH<sub>4</sub> (tetrahydrobiopterin); CGMP (cyclic guanosine monophosphate); GTP (guanosine triphosphate); eNOS (endothelial nitric oxide synthase); MAPK (mitogen-activated protein kinase); NADPH (nicotinamide adenine dinucleotide phosphatase); N(G)-dimethyl-L-arginine (endogenous NOS inhibitor); p- (phosphorylated protein); PTP (protein tyrosine phosphatase); Rec (receptor); sGC (soluble guanylyl cyclase); SHP-2 (SH2 domain-containing protein tyrosine phosphatase-2); VSMC (vascular smooth muscle cells). Taken from Touyz RM and Briones AM. Hypertension Research. 2011; 5-14<sup>10</sup>.

### Role of vascular factors and hormones

The NO has an important role as a paracrine regulator of vascular tone. NO, physiologically, inhibits leukocyte adhesion to the endothelium, migration and proliferation of VSMC, and platelet aggregation, thereby maintaining the vascular endothelium healthy. Consequently it has many beneficial effects. A decrease in the bioavailability of NO in the vasculature reduces its vasodilatory capacity and contributes to HT. The enzyme that catalyzes the formation of NO from O<sub>2</sub> and arginine is the NOS, which in fact is a whole family of enzymes. The eNOS is the predominant isoform of the NOS in the vascular wall. The stimulation of the receptors by its agonists leads to the rapid activation of the enzyme and the shear stress and the allosteric modulators are important regulators of its activity<sup>40</sup>. In addition to its vasodilatory and antiproliferative action, NO has an important role that antagonizes the effects of angiotensin II, endothelin and ROS. The NO diffuses as a gas to the underlying smooth muscle where it interacts with different molecular receptors such as soluble guanylyl cyclase.

The normal production of NO has a crucial role in maintaining the physiological conditions within the cardiovascular system. The L-arginine, a substrate for eNOS, appears to be a promising compound for the preservation of NO formation; however, the L-arginine fails to prevent high rates of blood pressure and left ventricular remodeling due to chronic treatment with the methyl ester of N-nitro-L-arginine which is an inhibitor of eNOS<sup>41</sup>. Captopril completely prevents HT by NO deficiency without improving NOS activity. The ON also exerts a negative feedback on ACE. The thiol groups protect NO from oxidation by collecting waste from ROS and forming nitrosothiols, these effects prolong the lifespan and duration of the NO action<sup>42,43</sup>.

The reduced levels of NO may be attributed to the increase in ROS. The O<sub>2</sub><sup>-</sup> combines with NO to form peroxynitrite which oxidizes BH<sub>4</sub> and destabilizes eNOS to produce more O<sub>2</sub><sup>-</sup><sup>33,34</sup>, which further increases the OS. Therefore, the balance between the NO and angiotensin II in the vasomotor center is very important for the regulation of sympathetic tone.

### Renin-Angiotensin system

This system has an important role in the development of cardiovascular disease. Angiotensin II is a potent vasoactive peptide that forms in the vascular bed rich

in ACE. When its production rises above normal levels, it induces vascular remodeling and endothelial dysfunction, which are associated with high rates of BP; and as it is a potent activator of NADPH oxidase, it contributes to the production of ROS<sup>44,45</sup>. In rats and mice where HT is induced by infusion of angiotensin II, the expression of the subunits of the NADPH oxidase, the oxidizing activity and ROS generation are increased<sup>46</sup>. This angiotensin II not only increases the activity of NADPH oxidase, but also upregulates the activity of SOD, possibly to compensate the increased levels of ROS. In situations where this compensatory effect is effective, the ROS levels may remain normal, even in pro-oxidant conditions. However, when ROS production becomes uncontrollable, the compensatory mechanisms are insufficient and physiopathologic consequences are triggered<sup>47</sup>.

Captopril and enalapril prevented the increase in the BP of young rats with induced HT by inhibiting ACE. Captopril, probably due to the antioxidant role of its thiol group, was more effective than enalapril in its antihypertensive effect<sup>48</sup>. However, the NO not only antagonizes the effects of angiotensin II on vascular tone, cell growth and renal sodium excretion, but also down-regulates the synthesis of ACE and the expression angiotensin I receptors. Therefore, the ACE inhibition upregulates the expression of the NOS. The ability of angiotensin II to induce endothelial dysfunction is also due to its ability to downregulate the soluble guanylyl cyclase, causing damage in the signaling of NO/cGMP.

### Acetylcholine

In blood vessels, the acetylcholine induces endothelial dilatation by the production of endothelial factors, mainly NO, which diffuses into the layer of underlying vascular smooth muscle and induces vasorelaxation. Decreased bioavailability of NO causes a significant reduction in vasodilation mediated by acetylcholine<sup>48,49</sup> and the result of the overall increase in ROS is a decrease in NO.

### ET-1

Endothelins are potent vasoconstrictors isopeptides produced by different vascular tissues, including the vascular endothelium. ET-1 is the main endothelin that is generated in the endothelium and the most im-

portant in the cardiovascular system. When administered in high concentrations, it behaves as a potent vasoconstrictor that is capable of exerting a number of effects, such as altering BP. The ET-1 acts through two receptors, ET<sub>A</sub> and ET<sub>B</sub>. ET<sub>A</sub> receptor mediates vasoconstriction via the activation of NADPH oxidase, xanthine oxidase, lipoxygenase, the uncoupling of eNOS and the enzymes of the mitochondrial respiratory chains. ET<sub>B</sub> receptor induces the relaxation of endothelial cells<sup>50</sup>. Many factors that normally stimulate the synthesis of ET-1, (e.g. thrombin and angiotensin II) also increase vasodilators such as prostacyclin (PGI<sub>2</sub>) and NO, or both, which oppose the vasoconstrictor function of ET-1. Hence it has been reported that essential HT is characterized by increased vasoconstrictor tone mediated by ET-1, in addition to a reduction of ET<sub>B</sub> due to the decreased production of NO, or its poor bioavailability.

### Urotensin II

Urotensin II is a powerful vasoactive peptide<sup>51</sup>, and is, in fact, the most potent vasoconstrictor that has been identified. It works through the activation of NADPH oxidase. The role of urotensin II in HT is still not well understood, because the vasoconstrictor response appears to be variable and highly dependent on the vascular bed; However, vasoconstriction is not the only effect, because urotensin II receptors have been identified in other organs<sup>52,53</sup>. This peptide also seems to function as a potent vasodilator in some isolated vessels<sup>54</sup>.

### Norepinephrine

The VSMC are primarily innervated by the sympathetic nervous system through the adrenergic receptors. Three types of receptors are present in VSMC:  $\alpha_1$ ,  $\alpha_2$  and  $\beta_2$ . Norepinephrine stimulates the proliferation of vascular smooth muscle. In addition, the overexpression of NO increases BP due to the activation of the sympathetic nervous system mediated by an increase in OS<sup>16</sup>.

### Prostaglandins

PGI<sub>2</sub> is another vasodilator produced in the endothelium which relaxes the vascular smooth muscle. It is released in higher amounts in response to different

compounds such as thrombin, arachidonic acid, histamine and serotonin. The enzyme prostaglandin H<sub>2</sub> synthase uses arachidonic acid as substrate to produce prostaglandin H<sub>2</sub>, which is converted into vasoactive molecules such as PGI<sub>2</sub>. The isoform of the enzyme prostaglandin H<sub>2</sub> synthase-2 may participate in the vascular disorder under conditions of OS; thus, peroxy-nitrite inhibits the enzymatic activity of PGI<sub>2</sub> synthetase and affects the vasodilation mediated by PGI<sub>2</sub>.

### Homocysteine

This molecule may have an important role in the pathogenesis of essential HT. An elevated homocysteinemia decreases the vasodilation of NO, increases OS, stimulates proliferation of the vascular smooth muscle and alters the elastic properties of the vessel wall. Homocysteine therefore contributes to the elevation of blood pressure. In addition, its high levels could cause oxidative damage to the endothelium<sup>55</sup>. Correction of the homocysteinemia by the administration of B<sub>6</sub>, B<sub>12</sub> and folic acid, may be a useful therapy in HT<sup>6</sup>; however, more controlled and randomized trials are needed to establish the efficacy of these therapeutic agents.

## KIDNEY AND CENTRAL NERVOUS SYSTEM

So far we have discussed the importance of ROS in blood vessels and their relationship with HT; however, it is also important to emphasize the evidence that hypertensive stimuli, such as high salt intake and angiotensin II, not only promote ROS production at this level, but also in the kidney and the central nervous system. In addition, each of these sites also contributes to HT, or to the adverse effects of this disease<sup>56</sup>.

### Importance of OS in the kidney

Evidence suggests that ROS play a key role in the pathophysiological processes of various kidney diseases, which are considered causes and consequences of HT. With respect to the glomerular changes, ROS produce glomerulopathic lipoproteins and other glomerular inflammatory lesions<sup>57</sup>. A recent study showed that some lipid conglomerates produce NADPH oxid-

ase activation and the production of ROS, which is an important molecular mechanism that stimulates homocysteine, which favors the oxidative damage to podocytes. This damage may represent an early phenomenon that starts glomerulosclerosis during hyperhomocysteinemia<sup>58</sup>. One of the underlying mechanisms of tubulointerstitial injury, mediated by ROS, is the exposure of tubular cells to LDL, which may cause alterations in the interstitium tubules due to the production of ROS by NADPH oxidase<sup>59</sup>. Angiotensin II not only has a pathogenic role in the progression of tubulointerstitial injury, but also in obstructive nephropathy<sup>60,61</sup>; it also activates NADPH oxidase and generates  $O_2^-$  causing hypertrophy of renal tubular cells<sup>62</sup>.

There are findings that suggest that a high-fat diet induces renal inflammation and elevation of BP via ROS in the HT of rats<sup>63</sup>. Additionally, the metabolic syndrome is a risk factor for chronic renal failure (CRF), independent of diabetes and HT, probably due to the influence of ROS. The onset and maintenance of renal damage may worsen metabolic syndrome and HT<sup>64</sup>.

Various mechanisms of OS are involved in the endothelial dysfunction of CRF<sup>65</sup>, where ROS are elevated and are associated with the vascular reactivity of the endothelium and systolic BP<sup>66</sup>. High levels of ROS and asymmetric dimethylarginine have been identified as new risk factors of endothelial dysfunction<sup>67</sup>. Furthermore, high levels of this dimethylarginine have been found in CRF, which are associated with an increase in the thickness of the vascular intima and media, and an increase in cardiovascular accidents<sup>68</sup>.

### **Importance of OS in the central nervous system**

Besides the kidneys and blood vessels, the sympathetic nervous system, which is in turn regulated by the central nervous system, is involved in the pathogenesis of HT<sup>69</sup>. Recent studies suggest that increased central sympathetic stimulation increases BP<sup>70</sup>. There is also evidence that increased ROS generation in the brain contributes to the neural mechanisms involved in the HT of rats<sup>71</sup>.

The rostral ventrolateral medulla is the largest vasomotor center, and is essential in the maintenance of basal vascular tone<sup>72,73</sup>. Some results show that an increase in the ROS at this level increases the vasomotor stimulation of spontaneous HT in rats and thus contributes to the neural mechanisms of HT through

the sympathetic nervous system activation<sup>72</sup>. The paraventricular nucleus of the hypothalamus is, perhaps, the one most strongly associated with the neural mechanisms of the HT associated with ROS<sup>74</sup>. There is evidence that other brain regions are also involved in this type of HT. These studies suggest that the increased production of intracellular  $O_2^-$  in the subfornical organ is critical in the development of HT induced by angiotensin II<sup>75</sup>.

### **CONCLUSIONS**

Numerous data confirm the importance of ROS in the control of vascular function, by means of the regulation of endothelial function and vascular tone through a strict control of the redox-sensitive signaling pathways. The ROS are mediators in most of the physiological vasoconstrictors that increase the concentration of intracellular calcium. The  $O_2^-$  reduces the bioavailability of NO and uncouples the eNOS, which in turn increases even more the concentrations of  $O_2^-$ . The uncontrolled production/degradation of ROS causes the OS, which results in cardiovascular, renal and neural damage, and is associated with the elevation of BP. Although oxidative damage is not the only cause of HT, it favors and increases the elevation of BP in the presence of other pro-hypertensive factors. Results from experimental studies, and in animals, indicate the role of OS in the pathogenesis of HT, possibly through the activation of oxidant enzymes, where the NADPH oxidases and the mitochondrial ones have an important role. From a clinical point of view, it is necessary that these data on the causal role of ROS in human HT remain under investigation.

### **REFERENCES**

1. Kakar P, Lip GY. Towards understanding the aetiology and pathophysiology of human hypertension: where are we now? *J Hum Hypertens*. 2006;20(11):833-6.
2. Viel EC, Lemarié CA, Benkirane K, Paradis P, Schiffrin EL. Immune regulation and vascular inflammation in genetic hypertension. *Am J Physiol Heart Circ Physiol*. 2010;298(3):938-44.



3. Vaziri ND, Rodríguez-Iturbe B. Mechanisms of disease: oxidative stress and inflammation in the pathogenesis of hypertension. *Nat Clin Pract Nephrol*. 2006;2(10):582-93.
4. Kagota S, Tada Y, Kubota Y, Nejime N, Yamaguchi Y, Nakamura K, *et al*. Peroxynitrite is involved in the dysfunction of vasorelaxation in SHR/NDmcr-cp rats, spontaneously hypertensive obese rats. *J Cardiovasc Pharmacol*. 2007;50(6): 677-85.
5. Venereo JR. Daño oxidativo, radicales libres y anti-oxidantes. *Rev Cubana Med Milit*. 2002;31(2):126-33.
6. Jerlich A, Pitt AR, Schaur RJ, Spickett CM. Pathway of phospholipid oxidation by HOCl in human LDL detected by LC-MS. *Free Radic Biol Med*. 2000; 28(5):673-82.
7. Lavi S, Yang EH, Prasad A, Mathew V, Barsness GW, Rihal CS, *et al*. The interaction between coronary endothelial dysfunction, local oxidative stress, and endogenous nitric oxide in humans. *Hypertension*. 2008;51(1):127-33.
8. Johnson F, Giulivi C. Superoxide dismutases and their impact upon human health. *Mol Aspects Med*. 2005;26(4-5):340-52.
9. Mendez JI, Nicholson WJ, Taylor WR. SOD isoforms and signaling in blood vessels: evidence for the importance of ROS compartmentalization. *Arterioscler Thromb Vasc Biol*. 2005;25(5):887-8.
10. Touyz RM, Briones AM. Reactive oxygen species and vascular biology: implications in human hypertension. *Hypertension Res*. 2011;34(1):5-14.
11. Nishino T, Okamoto K, Eger BT, Pai EF, Nishino T. Mammalian xanthine oxidoreductase - mechanism of transition from xanthine dehydrogenase to xanthine oxidase. *FEBS J*. 2008;275(13):3278-89.
12. Moens AL, Kass DA. Tetrahydrobiopterin and cardiovascular disease. *Arterioscler Thromb Vasc Biol*. 2006;26(11):2439-44.
13. Liu Y, Zhao H, Li H, Kalyanaraman B, Nicolosi AC, Gutterman DD. Mitochondrial sources of H<sub>2</sub>O<sub>2</sub> generation play a key role in flow-mediated dilation in human coronary resistance arteries. *Circ Res*. 2003;93(9):573-80.
14. DeLano FA, Parks DA, Ruedi JM, Babior BM, Schmid-Schönbein GW. Microvascular display of xanthine oxidase and NADPH oxidase in the spontaneously hypertensive rat. *Microcirculation* 2006;13(7):551-66.
15. Badimón L, Martínez-González J. Disfunción endotelial. *Rev Esp Cardiol*. 2006;6(Supl A):21-30.
16. Kimura S, Zhang GX, Nishiyama A, Shokoji T, Yao L, Fan YY, *et al*. Mitochondria-derived reactive oxygen species and vascular MAP kinases: comparison of angiotensin II and diazoxide. *Hypertension*. 2005; 45(3):438-44.
17. Hool LC, Corry B. Redox control of calcium channels: from mechanisms to therapeutic opportunities. *Antioxid Redox Signal*. 2007;9(4):409-35.
18. Yoshioka J, Schreiter ER, Lee RT. Role of thioredoxin in cell growth through interactions with signaling molecules. *Antioxid Redox Signal*. 2006;8(11):2143-51.
19. Redón J, Oliva MR, Tormos C, Giner V, Chaves J, Iradi A, *et al*. Antioxidant activities and oxidative stress byproducts in human hypertension. *Hypertension*. 2003;41(5):1096-101.
20. Tanito M, Nakamura H, Kwon YW, Teratani A, Matsutani H, Shioji K, *et al*. Enhanced oxidative stress and impaired thioredoxin expression in spontaneously hypertensive rats. *Antioxid Redox Signal*. 2004;6(1):89-97.
21. Touyz RM. Reactive oxygen species, vascular oxidative stress, and redox signaling in hypertension: what is the clinical significance? *Hypertension*. 2004;44(3):248-52.
22. Briones AM, Touyz RM. Oxidative stress and hypertension: current concepts. *Curr Hypertens Rep*. 2010;12(2):135-42.
23. Bengtsson SH, Gulluyan LM, Disting GJ, Drummond GR. Novel isoforms of NADPH oxidase in vascular physiology and pathophysiology. *Clin Exp Pharmacol Physiol*. 2003;30(11):849-54.
24. Rodrigo R, Prat H, Passalacqua W, Araya J, Guichard C, Bächler JP. Relationship between oxidative stress and essential hypertension. *Hypertens Res*. 2007; 30(12):1159-67.
25. Landmesser U, Dikalov S, Price SR, McCann L, Fukui T, Holland SM, *et al*. Oxidation of tetrahydrobiopterin leads to uncoupling of endothelial cell nitric oxide synthase in hypertension. *J Clin Invest*. 2003; 111(8):1201-9.
26. Gavazzi G, Banfi B, Deffert C, Fiette L, Schappi M, Herrmann F, *et al*. Decreased blood pressure in NOX1-deficient mice. *FEBS Lett*. 2006;580(2):497-504.
27. Touyz RM, Schiffrin EL. Increased generation of superoxide by angiotensin II in smooth muscle cells from resistance arteries of hypertensive patients:

- role of phospholipase D-dependent NAD(P)H oxidase-sensitive pathways. *J Hypertens*. 2001;19(7):1245-54.
28. Ghiadoni L, Magagna A, Versari D, Kardasz I, Huang Y, Taddei S, *et al*. Different effect of antihypertensive drugs on conduit artery endothelial function. *Hypertension*. 2003;41(6):1281-6.
29. Yoshida J, Yamamoto K, Mano T, Sakata Y, Nishikawa N, Nishio M, *et al*. AT1 receptor blocker added to ACE inhibitor provides benefits at advanced stage of hypertensive diastolic heart failure. *Hypertension*. 2004;43(3):686-91.
30. Fearheller DL, Brown MD, Park JY, Brinkley TE, Basu S, Hagberg JM, *et al*. Exercise training, NADPH oxidase p22phox gene polymorphisms, and hypertension. *Med Sci Sports Exerc*. 2009;41(7):1421-8.
31. Zou MH, Cohen R, Ullrich V. Peroxynitrite and vascular endothelial dysfunction in diabetes mellitus. *Endothelium*. 2004;11(2):89-97.
32. Lassègue B, Clempus RE. Vascular NAD(P)H oxidases: specific features, expression, and regulation. *Am J Physiol Regul Integr Comp Physiol*. 2003;285(2):277-97.
33. Kuzkaya N, Weissmann N, Harrison DG, Dikalov S. Interactions of peroxynitrite, tetrahydrobiopterin, ascorbic acid, and thiols: implications for uncoupling endothelial nitric-oxide synthase. *J Biol Chem*. 2003;278(25):22546-54.
34. Laursen JB, Somers M, Kurz S, McCann L, Warnholtz A, Freeman BA, *et al*. Endothelial regulation of vasomotion in apoE-deficient mice: implications for interactions between peroxynitrite and tetrahydrobiopterin. *Circulation*. 2001;103(9):1282-8.
35. Viel EC, Benkirane K, Javeshghani D, Touyz RM, Schiffrin EL. Xanthine oxidase and mitochondria contribute to vascular superoxide anion generation in DOCA-salt hypertensive rats. *Am J Physiol Heart Circ Physiol*. 2008;295(1):281-8.
36. Laakso JT, Teräväinen TL, Martelin E, Vaskonen T, Lapatto R. Renal xanthine oxidoreductase activity during development of hypertension in spontaneously hypertensive rats. *J Hypertens*. 2004;22(7):1333-40.
37. Han D, Antunes F, Canali R, Rettori D, Cadenas E. Voltage-dependent anion channels control the release of the superoxide anion from mitochondria to cytosol. *J Biol Chem*. 2003;278(8):5557-63.
38. Eto Y, Kang D, Hasegawa E, Takeshige K, Minakami S. Succinate-dependent lipid peroxidation and its prevention by reduced ubiquinone in beef heart submitochondrial particles. *Arch Biochem Biophys*. 1992;295(1):101-6.
39. Zhou L, Xiang W, Potts J, Floyd M, Sharan C, Yang H, *et al*. Reduction in extracellular superoxide dismutase activity in African-American patients with hypertension. *Free Radic Biol Med*. 2006;41(9):1384-91.
40. Michel JB, Feron O, Sase K, Prabhakar P, Michel T. Caveolin versus calmodulin. Counterbalancing allosteric modulators of endothelial nitric oxide synthase. *J Biol Chem*. 1997;272(41):25907-12.
41. Simko F, Luptak I, Matuskova J, Krajcovicova K, Sumbalova Z, Kucharska J, *et al*. L-arginine fails to protect against myocardial remodelling in L-NAME-induced hypertension. *Eur J Clin Invest*. 2005;35(6):362-8.
42. Zhang Y, Hogg N. S-Nitrosothiols: cellular formation and transport. *Free Radic Biol Med*. 2005;38(7):831-8.
43. Sládková M, Kojsová S, Jendeková L, Pechánová O. Chronic and acute effects of different antihypertensive drugs on femoral artery relaxation of L-NAME hypertensive rats. *Physiol Res*. 2007;56(Suppl 2):85-91.
44. Touyz RM. Reactive oxygen species and angiotensin II signaling in vascular cells – implications in cardiovascular disease. *Braz J Med Biol Res*. 2004;37(8):1263-73.
45. Hitomi H, Kiyomoto H, Nishiyama A. Angiotensin II and oxidative stress. *Curr Opin Cardiol*. 2007;22(4):311-5.
46. Landmesser U, Cai H, Dikalov S, McCann L, Hwang J, Jo H, *et al*. Role of p47(phox) in vascular oxidative stress and hypertension caused by angiotensin II. *Hypertension*. 2002;40(4):511-5.
47. Taniyama Y, Griendling KK. Reactive oxygen species in the vasculature: molecular and cellular mechanisms. *Hypertension*. 2003;42(6):1075-81.
48. Pechánová O. Contribution of captopril thiol group to the prevention of spontaneous hypertension. *Physiol Res*. 2007;56(Suppl 2):41-8.
49. Bitar MS, Wahid S, Mustafa S, Al-Saleh E, Dhaunsi GS, Al-Mulla F. Nitric oxide dynamics and endothelial dysfunction in type II model of genetic diabetes. *Eur J Pharmacol*. 2005;511(1):53-64.
50. Gomez-Alamillo C, Juncos LA, Cases A, Haas JA, Romero JC. Interactions between vasoconstrictors and vasodilators in regulating hemodynamics of

- distinct vascular beds. *Hypertension*. 2003;42(4):831-6.
51. Djordjevic T, BelAiba RS, Bonello S, Pfeilschifter J, Hess J, Görlach A. Human urotensin II is a novel activator of NADPH oxidase in human pulmonary artery smooth muscle cells. *Arterioscler Thromb Vasc Biol*. 2005;25(3):519-25.
  52. Matsushita M, Shichiri M, Imai T, Iwashina M, Tanaka H, Takasu N, et al. Co-expression of urotensin II and its receptor (GPR14) in human cardiovascular and renal tissues. *J Hypertens*. 2001;19(12):2185-90.
  53. Jégou S, Cartier D, Dubessy C, Gonzalez BJ, Chatenet D, Tostivint H, et al. Localization of the urotensin II receptor in the rat central nervous system. *J Comp Neurol*. 2006;495(1):21-36.
  54. Stirrat A, Gallagher M, Douglas SA, Ohlstein EH, Berry C, Kirk A, et al. Potent vasodilator responses to human urotensin-II in human pulmonary and abdominal resistance arteries. *Am J Physiol Heart Circ Physiol*. 2001;280(2):925-8.
  55. Rodrigo R, Passalacqua W, Araya J, Orellana M, Rivera G. Homocysteine and essential hypertension. *J Clin Pharmacol*. 2003;43(12):1299-306.
  56. Harrison DG, Gongora MC. Oxidative stress and hypertension. *Med Clin North Am*. 2009;93(3):621-35.
  57. Rodrigo R, Rivera G. Renal damage mediated by oxidative stress: a hypothesis of protective effects of red wine. *Free Radic Biol Med*. 2002;33(3):409-22.
  58. Zhang C, Hu JJ, Xia M, Boini KM, Brimson C, Li PL. Redox signaling via lipid raft clustering in homocysteine-induced injury of podocytes. *Biochim Biophys Acta*. 2010;1803(4):482-91.
  59. Piccoli C, Quarato G, D'Aprile A, Montemurno E, Scrima R, Ripoli M, et al. Native LDL-induced oxidative stress in human proximal tubular cells: multiple players involved. *J Cell Mol Med*. 2009;15(2):375-95.
  60. Klahr S. Urinary tract obstruction. *Semin Nephrol*. 2001;21(2):133-45.
  61. Grande MT, Pérez-Barriocanal F, López-Novoa JM. Role of inflammation in tubulo-interstitial damage associated to obstructive nephropathy. *J Inflamm (Lond)* [Internet]. 2010 [Citado 2013 Abr 13];22(7):19. Disponible en: <http://www.journal-inflammation.com/content/7/1/19>
  62. Sachse A, Wolf G. Angiotensin II-induced reactive oxygen species and the kidney. *J Am Soc Nephrol*. 2007;18(9):2439-46.
  63. Chung S, Park CW, Shin SJ, Lim JH, Chung HW, Youn DY, et al. Tempol or candesartan prevents high-fat diet-induced hypertension and renal damage in spontaneously hypertensive rats. *Nephrol Dial Transplant*. 2010;25(2):389-99.
  64. Guarnieri G, Zanetti M, Vinci P, Cattin MR, Pirulli A, Barazzoni R. Metabolic syndrome and chronic kidney disease. *J Ren Nutr*. 2010;20(Suppl 5):19-23.
  65. Malyszko J. Mechanism of endothelial dysfunction in chronic kidney disease. *Clin Chim Acta*. 2010;411(19-20):1412-20.
  66. Costa-Hong V, Bortolotto LA, Jorgetti V, Consolim-Colombo F, Krieger EM, Lima JJ. Oxidative stress and endothelial dysfunction in chronic kidney disease. *Arq Bras Cardiol*. 2009;92(5):381-6.
  67. Zoccali C, Bode-Böger S, Mallamaci F, Benedetto F, Tripepi G, Malatino L, et al. Plasma concentration of asymmetrical dimethylarginine and mortality in patients with end-stage renal disease: a prospective study. *Lancet* 2001;358(9299):2113-7.
  68. Nanayakkara PW, Teerlink T, Stehouwer CD, Allajlar D, Spijkerman A, Schalkwijk C, et al. Plasma asymmetric dimethylarginine (ADMA) concentration is independently associated with carotid intima-media thickness and plasma soluble vascular cell adhesion molecule-1 (sVCAM-1) concentration in patients with mild-to-moderate renal failure. *Kidney Int*. 2005;68(5):2230-6.
  69. Grassi G. Assessment of sympathetic cardiovascular drive in human hypertension: achievements and perspectives. *Hypertension*. 2009;54(4):690-7.
  70. Guyenet PG. The sympathetic control of blood pressure. *Nat Rev Neurosci*. 2006;7(5):335-46.
  71. Kishi T, Hirooka Y, Kimura Y, Ito K, Shimokawa H, Takeshita A. Increased reactive oxygen species in rostral ventrolateral medulla contribute to neural mechanisms of hypertension in stroke-prone spontaneously hypertensive rats. *Circulation*. 2004;109(19):2357-62.
  72. Hirooka Y, Sagara Y, Kishi T, Sunagawa K. Oxidative stress and central cardiovascular regulation. Pathogenesis of hypertension and therapeutic aspects. *Circ J*. 2010;74(5):827-35.
  73. Sved AF, Ito S, Sved JC. Brainstem mechanisms of hypertension: role of the rostral ventrolateral medulla. *Curr Hypertens Rep*. 2003;5(3):262-8.
  74. Oliveira-Sales EB, Nishi EE, Carillo BA, Boim MA,

Dolnikoff MS, Bergamaschi CT, *et al.* Oxidative stress in the sympathetic premotor neurons contributes to sympathetic activation in renovascular hypertension. *Am J Hypertens.* 2009;22(5):484-92.  
75. Zimmerman MC, Lazartigues E, Sharma RV, Davisson

RL. Hypertension caused by angiotensin II infusion involves increased superoxide production in the central nervous system. *Circ Res.* 2004;95(2):210-6.